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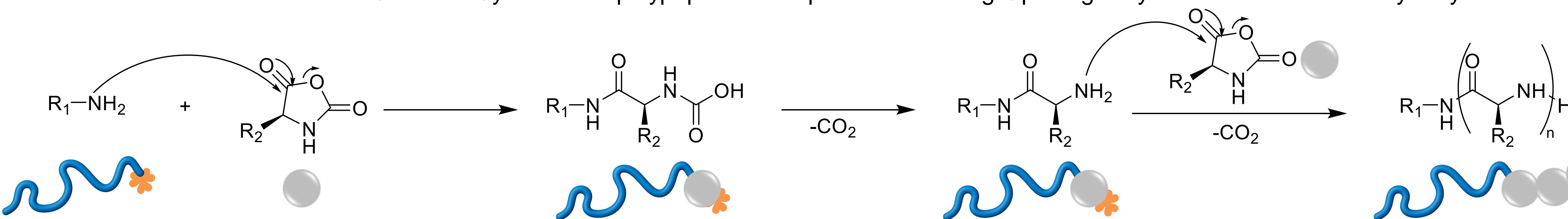
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Introduction Polypeptide-based nanoparticles have a wide range of applications, such as drug delivery, biosensing and bioimaging, due to their inherent biocompatibility and biodegradability.¹ These nanoparticles are typically synthesized from amphiphilic copolymers via the ring-opening polymerization (ROP) of *N*-carboxyanhydrides (NCAs).²

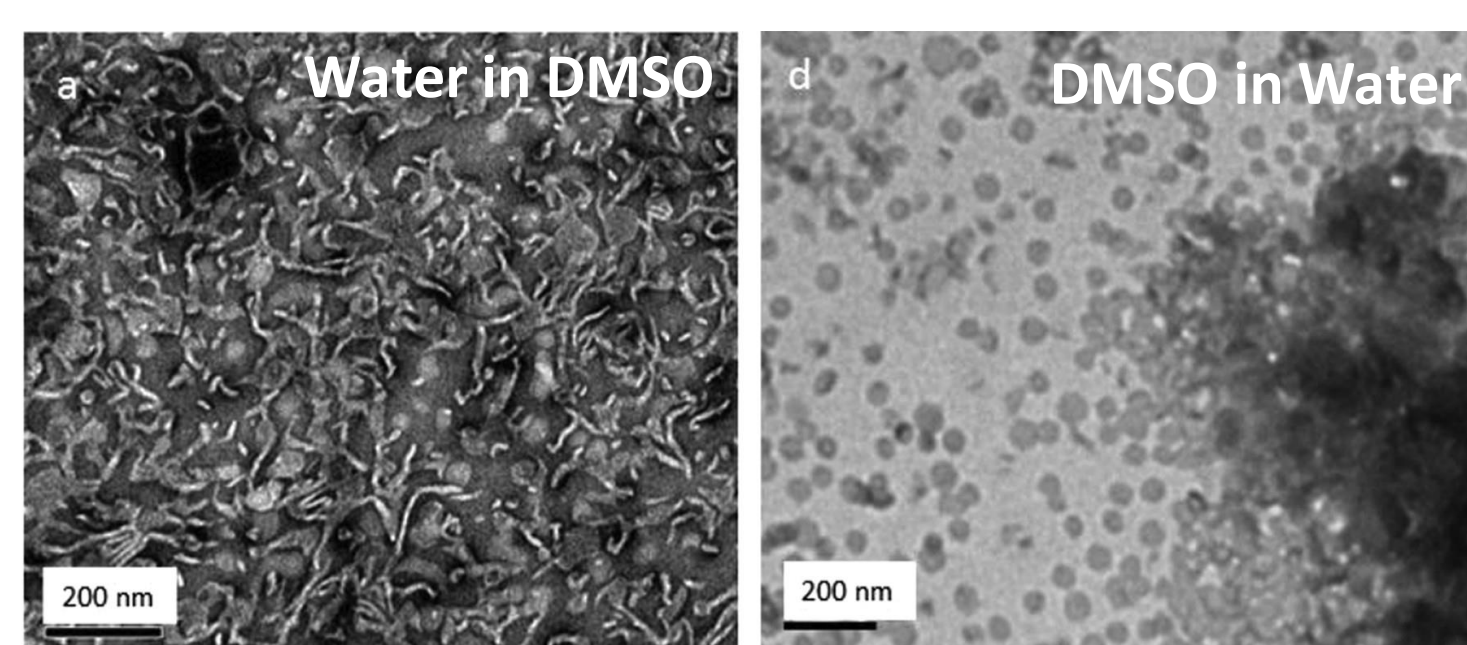
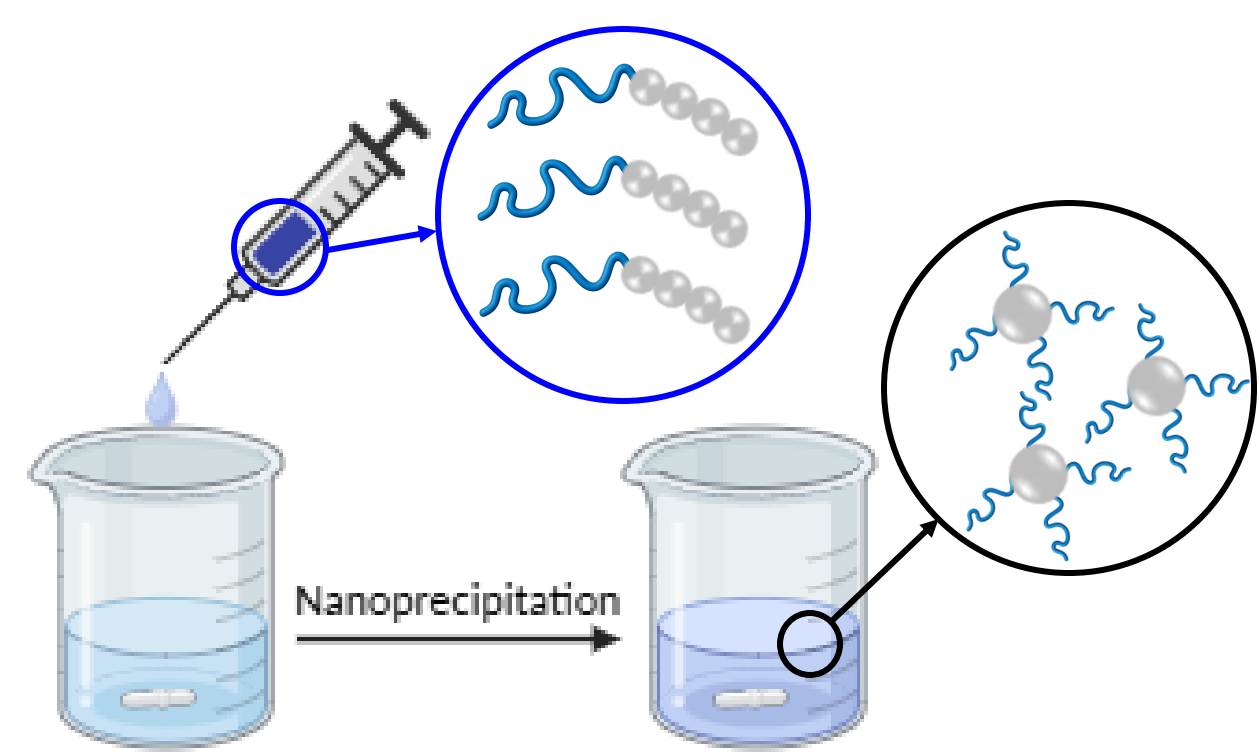
Schema: Synthesis of polypeptidic nanoparticles via Ring Opening Polymerization of *N*-carboxyanhydrides



R₁-Hydrophilic
R₂-Hydrophobic

- ✓ Scalable
- ✓ Economical method
- ✓ High molecular weight
- ✓ Polypeptide backbone
- ✓ Biodegradable and biocompatible

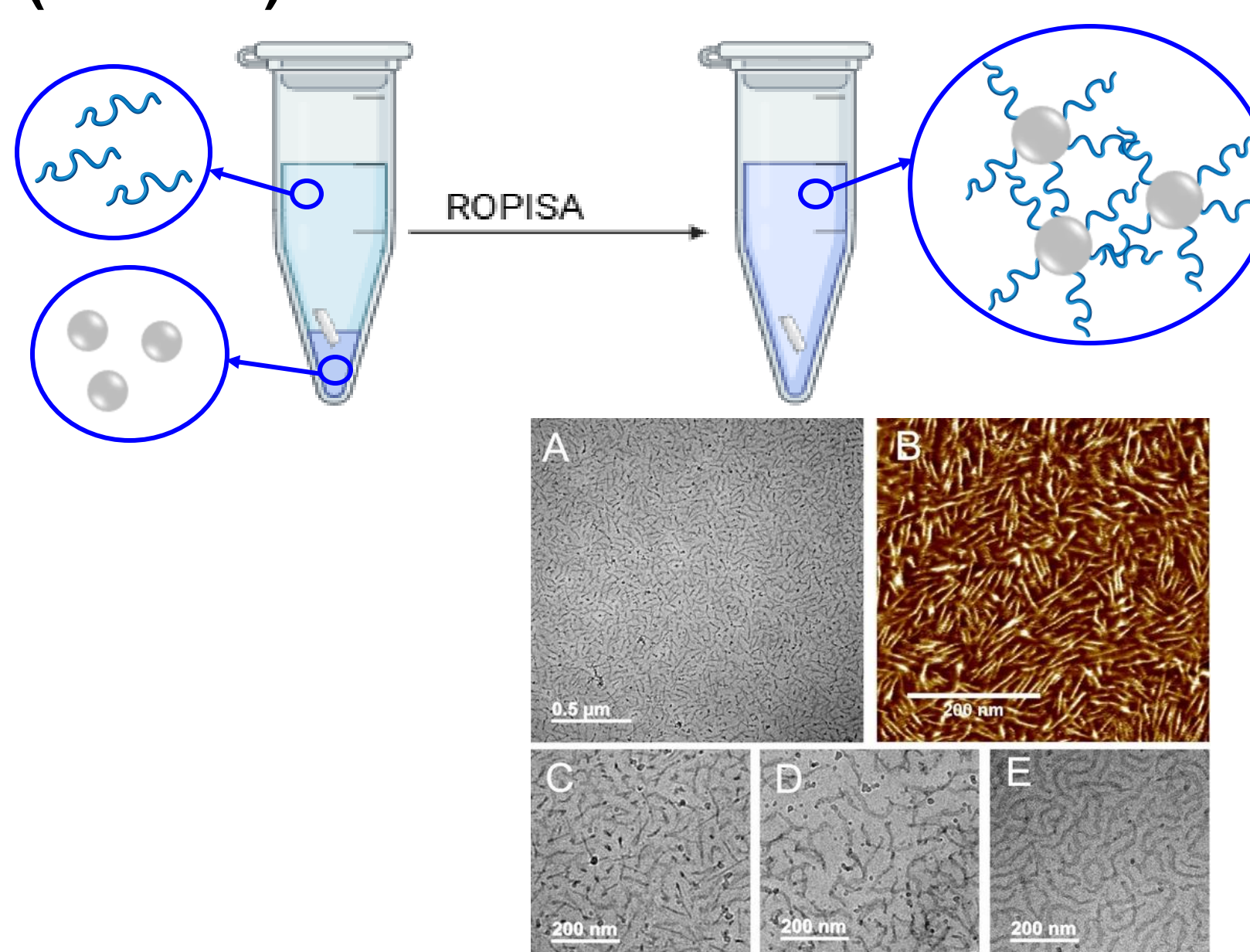
These nanoparticles are typically synthesized from amphiphilic copolymers via the **ring-opening polymerization (ROP)** of ***N*-carboxyanhydrides (NCAs)** in organic solvents, followed by a **nanoprecipitation** in water.³



PBLG₂₀-b-PGG₁₈ TEM images of samples obtained by instantaneously adding (a) water in DMSO and (d) DMSO in water³

- ✗ Use of organic solvents
- ✗ Hazardous up-scaling
- ✗ Stability of nanoparticles

A more promising approach involves the use of **Polymerization Induced by Self-Assembly (PISA)** combined with **Ring Opening Polymerization (ROP)** of ***N*-Carboxyanhydrides (ROPISA)**.⁴



- ✓ Reduces preparation steps
- ✓ Simplifies purification
- ✓ Increases the yield
- ✓ Formation of nanoobjects:
 - with a high solid content
 - with a biodegradable core
 - In H₂O, no solvent

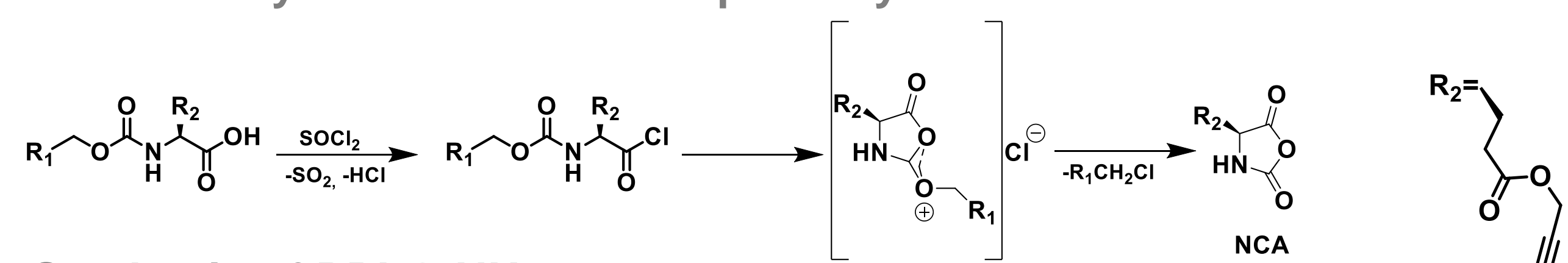
- ✗ No surface functionalization
- ✗ No biodegradable shell

Worm-like morphologies characterized by Cryo-TEM (A, C, D, E) and AFM (B).³

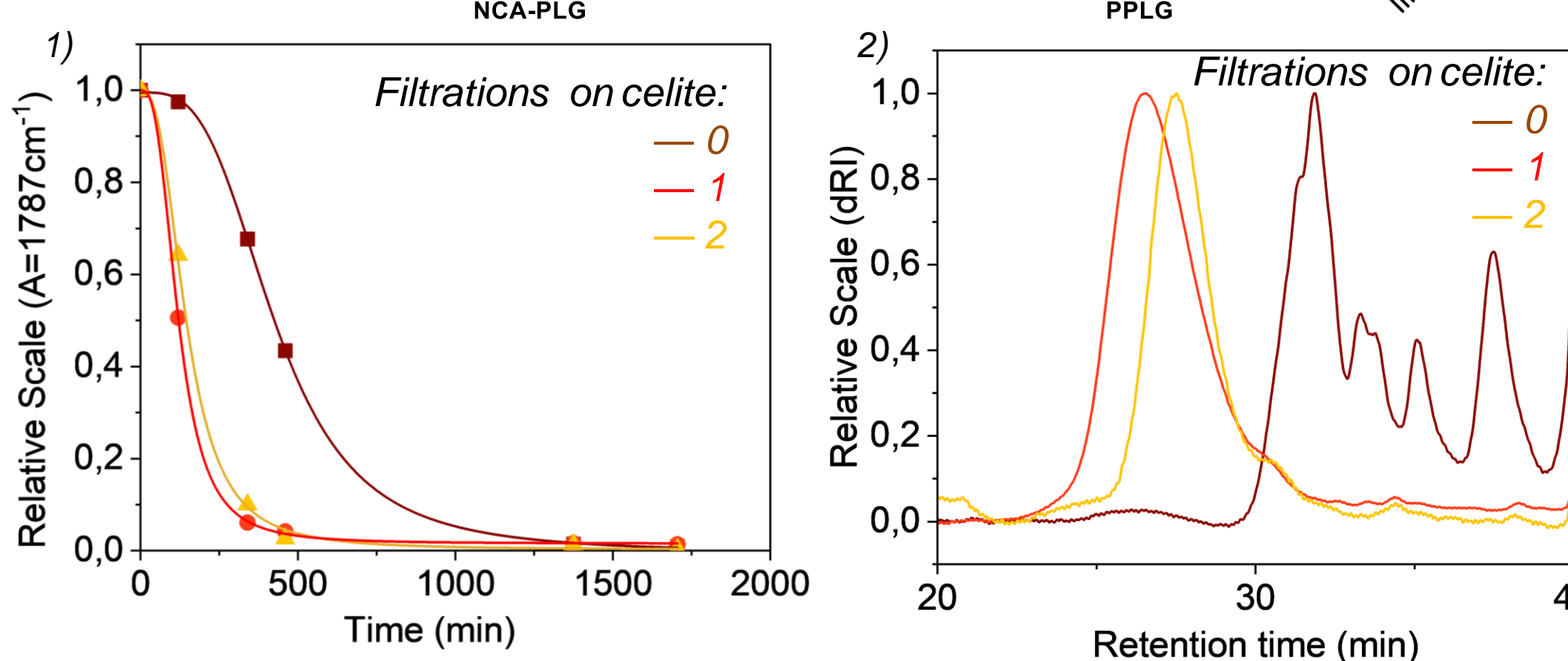
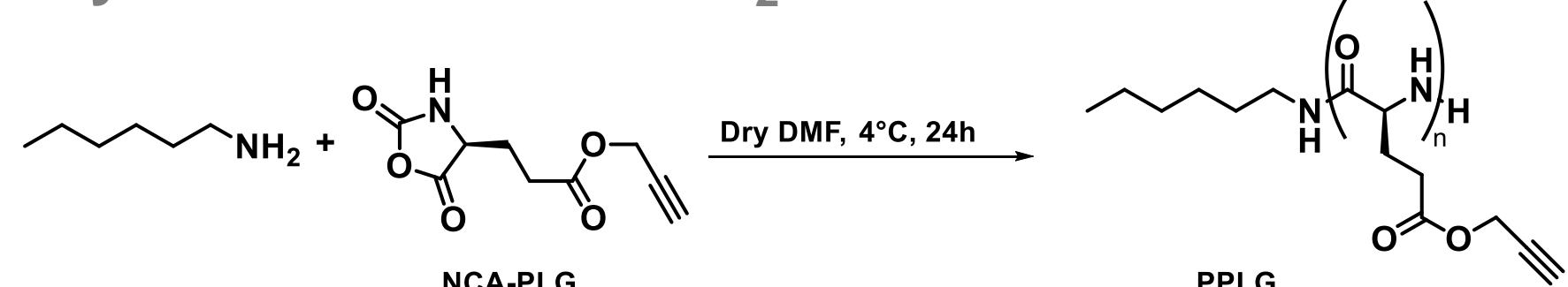
ROPISA with a hydrophobic clickable monomers: PLG-NCA

NCA clickable monomers offer a great **versatility** thanks to the dandling **alkyne groups** on the backbone which are availables for further **click chemistry**.

1. PLG-NCA synthesis via Leuchs pathway⁵



2. Synthesis of PPLG-NH₂



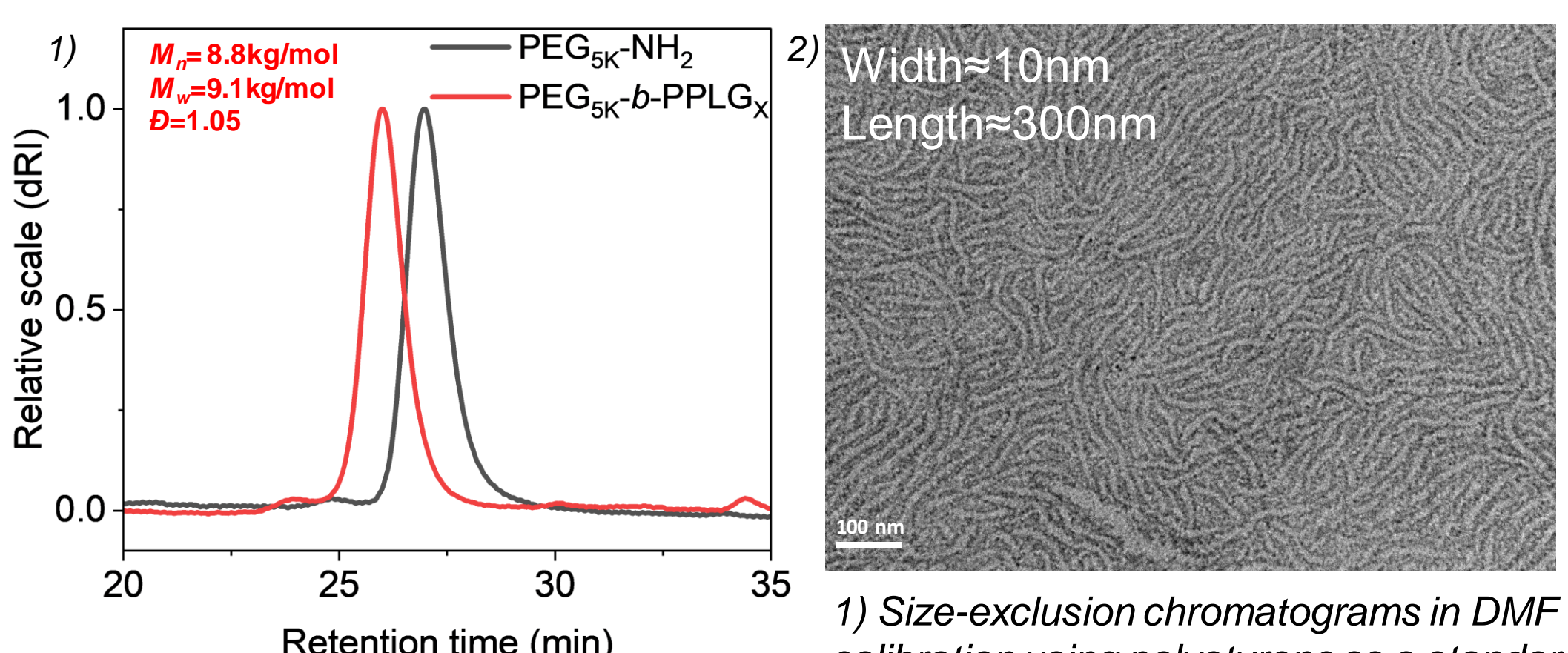
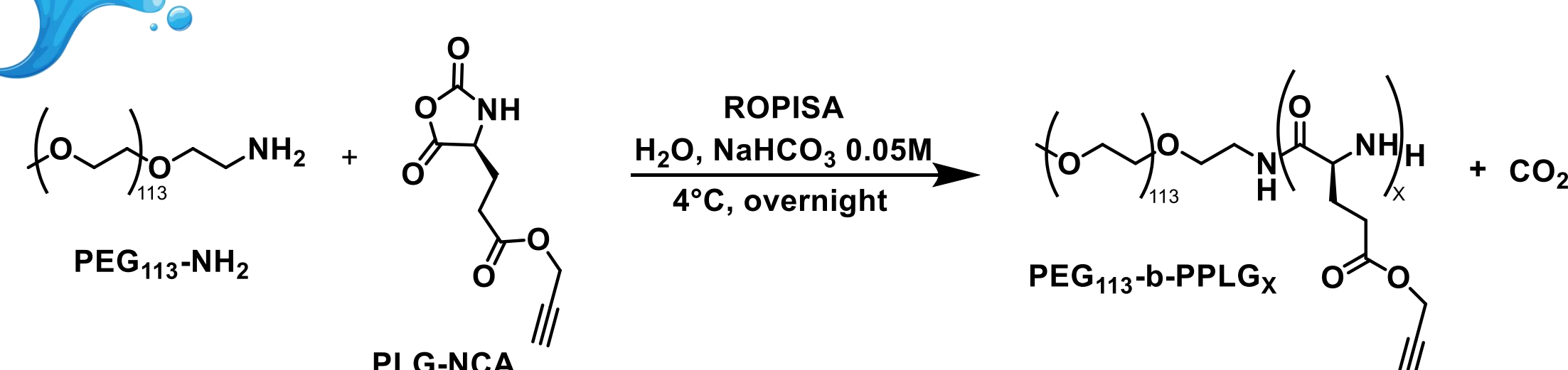
✓ Succesfull polymerization of PPLG via ROP of PLG-NCA in anhydrous conditions.

1) Kinetics by FITR at A=1787cm⁻¹ 2) Size-exclusion chromatograms in DMF (+LiBr 1mg.mL⁻¹), RI detection, calibration using polystyrene as a standard.

- Synthesis of PLG-NCA was done using 3 successive filtrations on celite, allowing to obtain a faster polymerization rates and better control of the polymerization.

Aqueous ROPISA of PLG-NCA using PEG-NH₂ as initiators

ROPISA of PLG-NCA



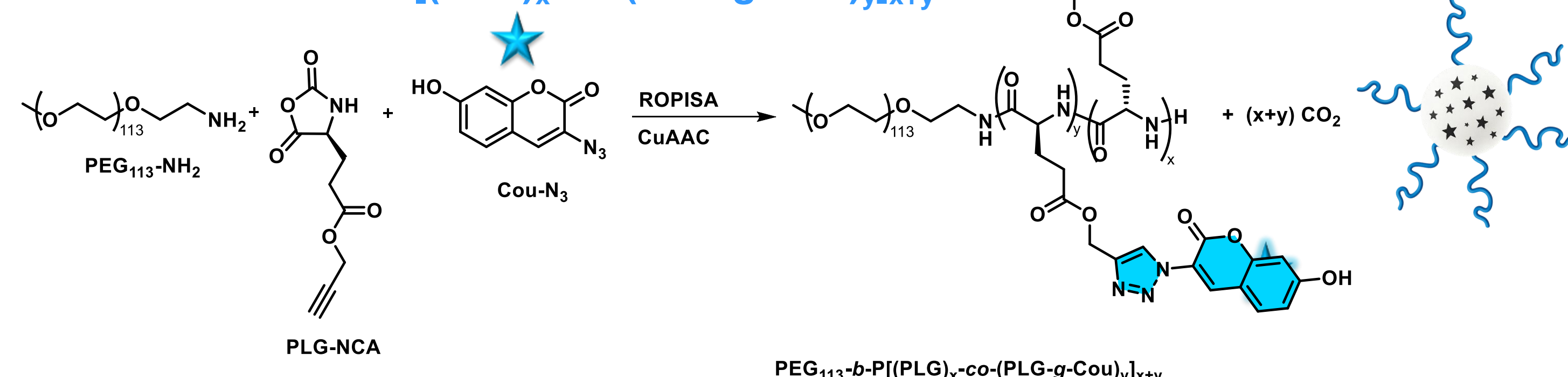
1) Size-exclusion chromatograms in DMF (+LiBr 1mg.mL⁻¹), RI detection, calibration using polystyrene as a standard. 2) Worm-like morphologies characterized by TEM

- ✓ succesfull synthesis of PPLG using ROPISA process.
- ✓ core-shell NPs of PEG-b-PPLG
- use of click chemistry to functionalize the PPLG.

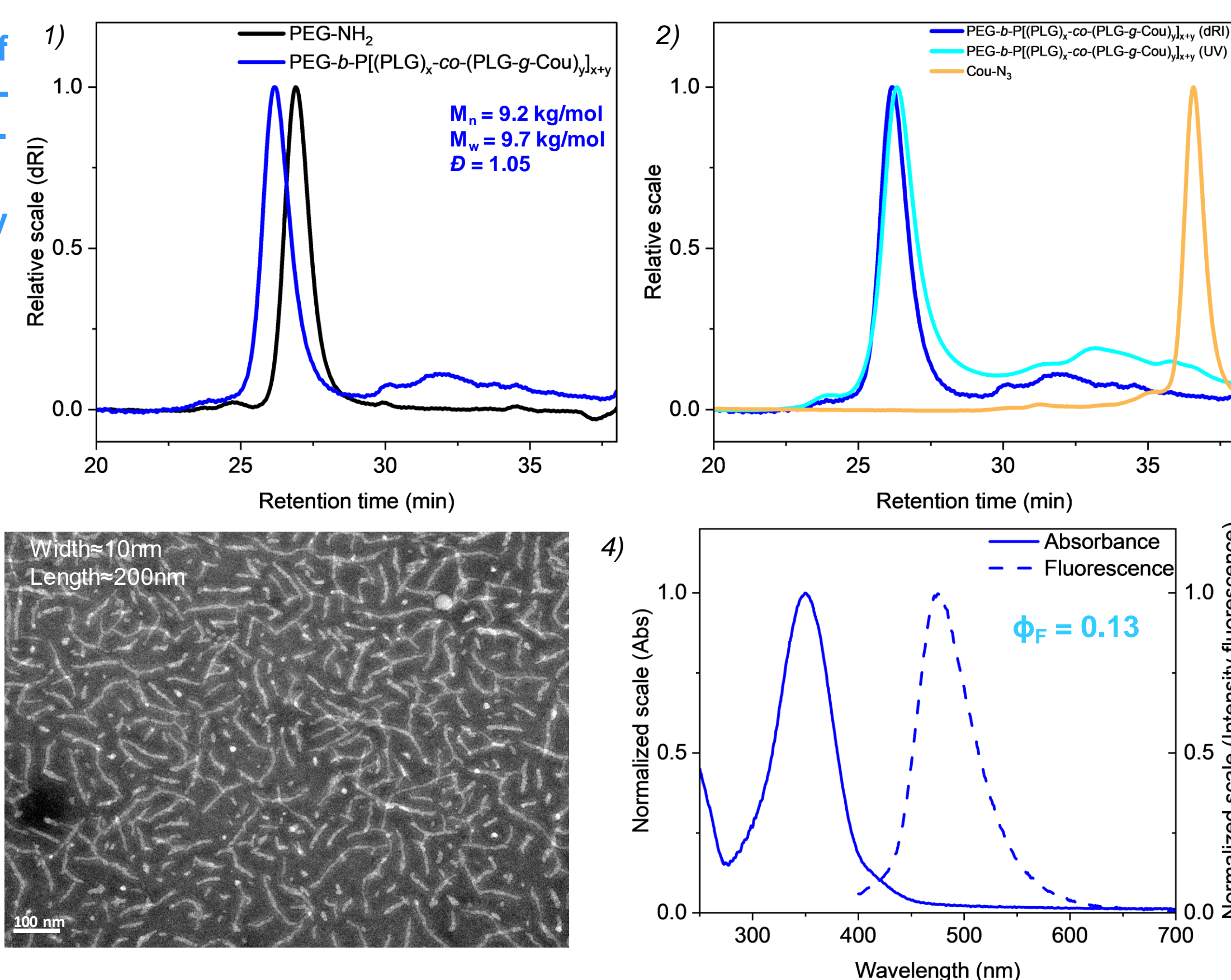
Click'n ROPISA: Synthesis of PEG-b-P[(PLG)_x-co-(PLG-g-Cou)]_{x+y}

Ring Opening Polymerization induced by Self Assembly of PLG-NCA using PEG-NH₂ as initiator and CuAAC⁶ of coumarine azide.

ROPISA of PEG-b-P[(PLG)_x-co-(PLG-g-Cou)]_{x+y}



- ✓ Succesfull synthesis of PEG-b-P[(PLG)_x-co-(PLG-g-Cou)]_{x+y} using ROPISA-CuAAC process.
- ✓ Nanoparticles morphology (Wormlike or spherics).
- ✓ Fluorescent.

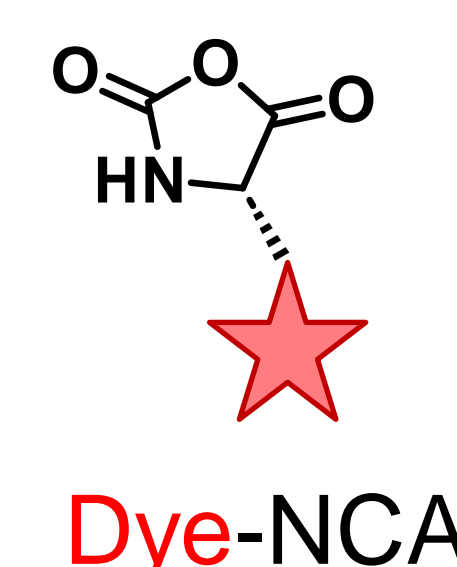


1) Size-exclusion chromatograms in DMF (+LiBr 1mg.mL⁻¹), RI detection, calibration using polystyrene as a standard. 2) Size-exclusion chromatograms in DMF (+LiBr 1mg.mL⁻¹), RI detection, UV detection calibration using polystyrene as a standard. 3) Worm-like and spherics (~15nm) nanoparticles characterized by TEM. 4) Absorbance and fluorescence emission spectra of PEG-b-P[(PLG)_x-co-(PLG-g-Cou)]_{x+y} in water at λ_{ex} = 350 nm.

Conclusions and perspectives

- ✓ Synthesis and polymerization of PLG-NCA
- ✓ ROPISA of PLG-NCA to obtain nanoparticles of PEG-b-PPLG
- ✓ ROPISA-CuAAC synthesis to achieve fluorescent core-shell nanoparticles in water
- ✓ Anisotropic objects

- Improvement of ROPISA and CuAAC synthesis conditions
- Synthesis of a library of nanoparticles using others dye-N₃
- Direct polymerization of new dye-NCA



References (1) Rasines Mazo, A.; *et al. Chem. Soc. Rev.* **2020**, 49 (14), 4737–4834. (2) Hadjichristidis N.; *Chem. Rev.* **2009**, 109, 5528–5578 (3) Huang J.; *et al. J. Am. Chem. Soc.*, 134, 119–122 (4) Gazon C.; *et al. Angewandte Chemie International Edition* **2020**, 59 (2), 622–626. (5) Leuchs, H.; *Berichte Dtsch. Chem. Ges.* **1906**, 39 (1), 857–861. (6) Huisgen R.; *Proc. Chem. Soc.*, **1961**, 357-396